New Cholinesterase-Inhibiting Steroidal Alkaloids from Sarcococca saligna

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Seven new steroidal alkaloids, 2-hydroxysalignarine-E (=(2′E,20S)-20-(dimethylamino)-2 β -hydroxy-3 β -(tigloylamino)pregn-4-ene; 1), 5,6-dihydrosarconidine (=(20S)-20-(dimethylamino)-3 β -(methylamino)-5 α -pregn-16-ene; 2), salignamine (=(20S)-20-(methylamino)-3 β -methoxypregna-5,16-diene; 3), 2-hydroxysalignamine (=(20S)-20-(dimethylamino)-2 β -hydroxy-3 β -methoxypregna-5,16-diene; 4), salignarine-F (=(2′E,20S)-20-(dimethylamino)-4 β -hydroxy-3 β -(tigloylamino)pregn-5-ene; 5), salonine-C (=(2′E,20S)-20-(dimethylamino)-3 β -(tigloylamino)pregna-4,14-diene; 6), and N-[formyl(methyl)amino]salonine-B (=(20S)-20-[formyl(methyl)amino]-3 β -methoxypregna-5,16-diene; 7) have been isolated from the MeOH extract of *Sarcococca saligna*, along with the six known alkaloids dictyophlebine (8), epipachysamine-D (9), saracosine (10), iso-N-formylchonemorphine (11), sarcodinine (12), and alkaloid-C (13). The structures of 1–7 were deduced from spectral data. Compounds 1–13 demonstrated significant activity against acetyl- and butyrylcholinesterase.

Introduction. – *Alzheimer*'s disease is the fourth leading cause of death in people over 65 years of age in industrialized countries [1]. A number of studies have identified a link between the onset of the disease and the enzymes acetylcholinesterase (AChE) and butyrylcholinesterase (BChE). As part of our efforts to identify new natural cholinesterase inhibitors, the whole plant of *Sarcococca saligna* was investigated, as its crude alkaloidal fraction showed significant activity in an *in vitro* high-throughput cholinesterase-inhibition assay.

The genus Sarcococca belongs to the Buxaceae family, with 16-20 species. It is predominantly distributed in South Asia, extending from Afghanistan through the Himalayas [2]. Some alkaloids of Sarcococca exhibit anticholinesterase and ganglion-blocking activities, while others potentiate the action of naturally secreted acetylcholine in the isolated diaphragm of rats and in the serum of rabbits [3]. Previous studies on this genus have led to the isolation of a number of pregnane-type steroidal alkaloids [4-15], including fifteen new cholinesterase-inhibiting alkaloids from our laboratory [15]. Here, we report the structures of the new compounds 1-7 and those of the known constituents 8-13 isolated from Sarcococca saligna.

Results and Discussion. – 2-Hydroxysalignarine-E (**1**), a colorless amorphous compound, showed a molecular ion (M^+) at m/z 442.3489 ($C_{28}H_{46}N_2O_2$; calc.: 442.3559) corresponding to seven degrees of unsaturation. Comparison of its IR, ¹H-NMR, and ¹³C-NMR data ($Tables\ 1$ and 2) with those of the known compound, salignenamide-B [11], suggested a pregnane-type steroidal skeleton, lacking one C=C bond. The presence of a tigloyl (E)-2-methylbut-2-enoyl) moiety in **1** ($\delta_{\rm H}$ 6.51, 1.77, 1.88; $\delta_{\rm C}$ 130.3,

$$R^{1} = \begin{pmatrix} 21 & R^{5} & R^{4} \\ 12 & 14 & 16 \\ 10 & H & (H) & 15 \\ R^{2} & 4 & 16 \\ R^{3} & (H) & 6 \end{pmatrix}$$

$$R^{1} = \begin{pmatrix} 11 & R^{5} & R^{4} & R^{5} \\ R^{4} & R^{4} & R^{4} & R^{5} \\ R^{2} & 4 & 16 \\ R^{3} & (H) & 6 \end{pmatrix}$$

$$Tigloylamino = \begin{pmatrix} 0 & 11 & 11 & 11 \\ 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11 & 11 & 11 & 11 \\ 11$$

R ¹	R^2	\mathbb{R}^3	R^4	R^5	Unsaturation
1 OH	tigloylamino	Н	Me	Me	$\Delta^{4,5}$
2 H	MeHN	Н	Me	Me	$\Delta^{16,17}$
3 H	MeO	Н	Me	Н	$\Delta^{5,6}$, $\Delta^{16,17}$
4 OH	MeO	Н	Me	Me	$\Delta^{5,6}, \Delta^{16,17}$
5 H	tigloylamino	OH	Me	Me	$\Delta^{5,6}$
6 H	tigloylamino	Н	Me	Me	$\Delta^{4,5}, \Delta^{14,15}$
7 H	MeO	Н	HCO	Me	$\Delta^{5,6}$, $\Delta^{16,17}$
8 H	MeHN	Н	Me	Me	_
9 H	PhCONH	Н	Me	Me	_
10 H	Me ₂ N	Н	Ac	Me	$\Delta^{5,6}$
11 H	Me ₂ N	Н	Н	HCO	_
12 H	Me ₂ N	Н	Me	Me	$\Delta^{5,6}$
13 H	MeO	Н	Me	Me	$\Delta^{5,6}$

Fig. 1. Structures of compounds 1-13 isolated from Sarcococca saligna. Note that C=C bonds (positions) are not explicitly shown in the steroidal framework, but indicated by $\Delta^{m,n}$. The tigloyl (=2-methylbut-2-enoyl) residue is numbered arbitrarily

13.9, 12.5) was consistent with literature values. In its $^1\text{H-NMR}$ spectrum, signals for two Me groups at quaternary (δ_{H} 0.84, 1.27), and one Me group at a secondary C-atom(s) (δ_{H} 1.07), a methine H-atom (δ_{H} 3.91) next to an OH group, and an olefinic H-atom (δ_{H} 5.51) were also clearly observed. In the COSY spectrum, the signals of both H–C(2) (δ_{H} 3.91) and H–C(4) (δ_{H} 5.51) were found to be coupled with H–C(3) (δ_{H} 4.19). The H–C(3) methine atom also showed a COSY cross-peak with the NH d (δ_{H} 6.70). The configuration at C(2) was inferred on the basis of the coupling constant ($W_{1/2}$ = 5.6 Hz) and the ROESY interactions (*Fig.* 2) between H–C(3) and H–C(2), suggesting the β -orientation of the C(3) tigloylamino and C(2)–OH groups. Thus, the structure of **1** was assigned as (2'*E*,20*S*)-20-(dimethylamino)-2 β -hydroxy-3 β -(tigloylamino)pregn-4-ene.

5,6-Dihydrosarconidine (2) was isolated as a white powder. HR-EI-MS Analysis showed the M^+ signal at m/z 358.3193 ($C_{24}H_{42}N_2^+$; calc.: 358.3198), with five degrees of unsaturation. In its 1H -NMR spectrum, two angular Me groups appeared at δ_H 0.84 (Me(18)) and 0.87 (Me(19)) (*Fig. 3*). A d at δ_H (1.13 (J=6.5 Hz) was assigned to Me(21), while two downfield s at δ_H 2.14 and 2.23 were assigned to HNMe and NMe $_2$, respectively. A q at δ_H 2.68 was assigned to H $_2$ C(20), indicating the presence of a C $_2$ C bond in ring D between C(16) and C(17). The base peak at m/z 343 was due to [M-Me] $_1$, and the lower abundance of the ion at M/z 72 (which is normally a base peak in such alkaloids) indicated a C $_2$ C bond between C(16) and C(17). Comparison of the

	1	2	3	4	5	6	7
H-C(1)	1.15, 1.85 (m)	1.90, 1.35 (m)	1.35, 2.20 (m)	1.15,2.20 (m)	1.85, 1.23 (m)	1.69, 1.15 (m)	1.35, 1.90 (m)
H-C(2)	$3.91 (m, W_{1/2} = 5.6)$	1.80, 1.75 (m)	1.80, 1.75 (m)	4.14 (br. s)	1.6, 1.21 (<i>m</i>)	2.6, 1.94 (m)	1.75, 1.80 (m)
H-C(3)	4.19 (m)	2.68 (m)	3.04 (m)	3.11 (br. s)	3.95 (br. s)	4.15 (m)	3.03 $(m, W_{1/2} = 17.7)$
H-C(4)	5.51 (br. s)	2.30, 2.60 (m)	2.11, 2.55 (m)	2.15, 2.55 (m)	3.99 (br. s)	5.78 (br. s)	2.14, 2.39 (m)
H-C(5)	_ ` `	1.65	_	_	_ ` `	_ ` `	_
H-C(6)	1.65, 2.11 (m)	1.32, 1.75 (m)	5.36 (br. s)	5.39 (br. s)	5.78 (br. s)	1.49, 2.15 (m)	5.34 (br. s)
H-C(7)	1.32, 1.84 (m)	2.00, 2.61 (m)	2.00, 2.60 (m)	2.00, 2.60 (m)	2.05, 2.5 (m)	1.88, 1.34 (m)	2.30, 1.5 (m)
H-C(8)	1.82	2.40	2.38	1.65	1.55	1.55	1.45
H-C(9)	0.75	1.38	1.27	1.00	1.23	0.88	1.35
H-C(10)	_	_	_	_	_	_	_
H-C(11)	1.61, 1.42 (m)	1.79, 1.99 (m)	1.20, 1.60 (m)	1.20, 1.60 (m)	2.13, 1.41 (<i>m</i>)	2.12, 1.39 (m)	1.7, 1.99 (m)
H-C(12)	1.36, 1.84 (m)	1.58, 1.50 (m)	$1.40, 1.70 \ (m)$	$1.40, 1.70 \ (m)$	1.52, 1.20 (m)	1.35, 1.23 (m)	1.53, 1.22 (m)
H-C(13)	_	_	_	_	_	_	_
H-C(14)	1.36	2.45	2.14	1.35	1.32	_	1.75
H-C(15)	2.01, 1.88 (m)	1.85, 2.06 (m)	1.65, 2.10 (m)	1.70, 2.10 (m)	1.7, 1.92 (m)	5.41 (br. s)	1.6, 1.20 (m)
H-C(16)	1.59, 1.61 (m)	5.55 (br. s)	5.55 (br. s)	5.64 (br.s)	1.63, 1.73 (m)	2.06, 1.65 (m)	5.78 (br. s)
H-C(17)	1.25	_ ` ´	_ ` ´	_ ` `	1.2	1.1	= ` ´
H-C(18)	0.84(s)	0.84(s)	0.84(s)	0.86(s)	0.82(s)	0.86(s)	0.75(s)
H-C(19)	1.27(s)	0.87(s)	1.03 (s)	1.21 (s)	1.04(s)	1.06(s)	1.01 (s)
H-C(20)	2.80(m)	2.68 (q, J = 6.6)	3.08 (d, J = 6.6)	2.90 (q, J = 6.6)	2.80(s)	2.82(m)	4.19 (m)
H-C(21)	1.07 (d, J = 6.6)	1.13 (d, J = 6.5)	1.17 (d, J = 6.5)	1.14 (d, J = 6.5)	1.38 (d, J = 6.5)	1.73 (d, J = 6.5)	1.3 (d, J = 6.5)
C(3)-NMe	_	2.14 (s)	_	_	_	_	_
C(20)-NMe	2.20(s)	2.23(s)	2.33(s)	2.28(s)	2.23(s)	2.91(s)	2.64/2.72 ^a)
H-C(1)	_	-	-	-	_	_	_ ′
H-C(2')	_	_	_	_	_	_	_
H-C(3')	6.51 (q, J = 6.9)	_	_	_	6.34 (q, J = 6.9)	6.34 (q, J = 6.9)	_
H-C(4')	1.77 (d, J = 6.9)	_	_	_	1.73 (d, J = 6.9)	1.78 (d, J = 6.9)	_
;;					1.7	1.1	

3.38(s)

3.34(s)

Table 1. ${}^{1}H$ -NMR Data of the New Steroidal Alkaloids $\mathbf{1}$ - $\mathbf{7}$ ($\delta_{\rm H}$ in ppm, J in Hz; solvent: CDCl₃)

1.88 (s)

H-C(5')

HCO

MeO

8.02/8.19^a)

3.33(s)

1.82 (s)

1.83 (s)

^a) Signal splitting due to rotational isomerism of the amido group.

Table 2. ¹³C-NMR Data of the New Steroidal Alkaloids **1–7** (δ_C in ppm; solvent: CDCl₃)

	1	2	3	4	5	6	7
C(1)	33.3	34.3	42.4	42.4	34.5	34.5	34.3
C(2)	69.6	30.5	29.6	67.0	36.5	37.4	29.6
C(3)	50.7	53.4	79.2	81.2	45.2	45.5	81.8
C(4)	115.3	35.0	32.3	32.4	68.8	126.4	32.5
C(5)	151.8	42.5	139.0	141.0	140.5	149.4	141.1
C(6)	41.8	27.6	121.2	121.2	129.1	30.3	121.3
C(7)	34.4	32.7	31.8	31.8	30.2	31.6	33.3
C(8)	32.8	34.1	33.1	30.0	35.5	33.5	32.3
C(9)	54.5	55.7	53.7	51.4	54.5	57.3	49.2
C(10)	38.1	35.4	35.5	37.0	39.5	39.5	35.5
C(11)	21.3	20.4	20.6	20.6	20.8	20.4	20.9
C(12)	34.4	31.9	34.6	34.6	31.8	34.3	31.9
C(13)	46.8	46.5	45.3	47.3	45.6	46.8	47.5
C(14)	56.9	57.2	54.7	56.7	55.5	154.6	56.4
C(15)	31.2	31.2	30.2	31.3	34.4	123.3	31.2
C(16)	118.7	123.3	130.3	130.3	32.7	31.3	125.8
C(17)	151.8	156.0	142.3	149.3	51.5	51.5	153.4
C(18)	15.9	12.7	19.7	19.7	15.8	15.9	18.3
C(19)	19.4	16.9	20.2	20.2	18.7	18.8	19.2
C(20)	59.2	59.3	61.7	61.7	57.9	59.5	57.3
C(21)	16.0	15.8	14.0	16.0	19.3	19.7	15.3
C(3)-NMe	_	30.3			_	-	-
C(20)-NMe	42.3	42.3	42.3	42.3	42.5	42.5	22.3/21.9 ^a)
C(1')	169.0	-		-	168.3	168.3	-
C(2')	132.1	_	_	_	131.3	131.5	_
C(3')	130.3	_	_	_	130.8	130.3	_
C(4')	13.9	-		-	11.5	12.3	-
C(5')	12.5	_	_	_	13.1	13.9	_
MeO	_		55.2	55.9	_		55.5
HCO	-	-	-	-	-	-	168.6

^a) Signal splitting due to rotational isomerism of the amido group.

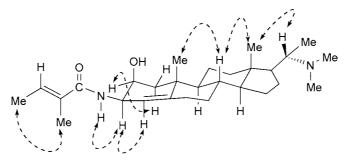


Fig. 2. Selected ROESY interactions in 2-hydroxysalignarine-E (1)

UV, IR, and ¹H-NMR data of **2** with those of the known alkaloid sarconidine [4] indicated close structural similarity, except for the lack of the C(4)=C(5) bond in **2**. Thus, the structure of compound **2** was assigned as (20S)-20-(dimethylamino)-3 β -(methylamino)-5 α -pregn-16-ene.

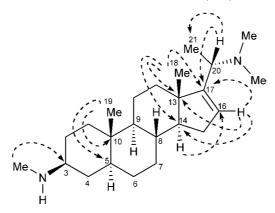


Fig. 3. Selected HMBC correlations in 5,6-dihydrosarconidine (2)

Salignamine (3) has a molecular formula of $C_{23}H_{37}NO$, as deduced by HR-EI-MS (m/z 343.2880 for M^+). The 1H - and ^{13}C -NMR spectra of 3 ($Tables\ 1$ and 2) were comparable to those of the known alkaloid-C (13), which was also isolated during this study, with an additional olefinic H-atom at $\delta_{\rm H}$ 5.55, and a downfield δ -shifted Me signal at $\delta_{\rm H}$ 2.33 in 3. The EI mass spectrum of 3 indicated that the extra double bond was between C(16) and C(17) (lower abundance of H_3C -HC=N⁺HMe at m/z 58, generally a base peak in HNMe-containing steroidal alkaloids with a saturated ring D [17]). The MeO group in 3 was placed at C(3) based on comparison of its 1H - and 13 C-NMR data with those of alkaloid-C (13) [5]. This was further supported by a HMBC experiment ($Fig.\ 4$), in which a correlation was observed between H–C(3) ($\delta_{\rm H}$ 3.04) and C(4) ($\delta_{\rm C}$ 32.3), which, in turn, were coupled with C(5) ($\delta_{\rm C}$ 139.0). Consequently compound 3 was deduced as (20S)-20-(methylamino)-3 β -methoxypregna-5,16-diene.

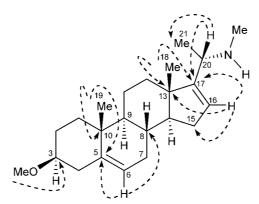


Fig. 4. Selected HMBC correlations in salignamine (3)

2-Hydroxysalignamine (4) had a molecular formula of $C_{24}H_{39}NO_2$ (m/z 373.2982; calc.: 373.2980), and its mass-fragmentation pattern, IR, and 1H - and 1S C-NMR spectra (*Tables 1* and 2) were distinctly similar to those of compound 13, with an additional OH group (IR: 3425 cm $^{-1}$). The location of the C(2)-OH group in 4 was assigned by a

HMBC experiment, in which correlations were observed between H–C(2) ($\delta_{\rm H}$ 4.14) and C(3) ($\delta_{\rm C}$ 81.2), and between H–C(3) ($\delta_{\rm H}$ 3.11) and C(5) ($\delta_{\rm C}$ 141.0). The configuration at C(2) was assigned as β on the basis of the coupling constant ($W_{1/2}$ = 5.5 Hz) and ROESY interaction (*Fig.* 5) between H–C(1a) and H–C(2), indicating that both were α-oriented. Thus, compound **4** was assigned as (2'E,20S)-20-(dimethyl-amino)-2 β -hydroxy-3 β -methoxypregna-5,16-diene.

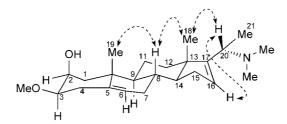


Fig. 5. Selected ROESY interactions in 2-hydroxysalignamine (4)

Salignarine-F (**5**) was obtained as a yellow gum. Its HR-EI mass spectrum displayed M^+ at 442.3577 ($C_{28}H_{46}N_2O_2^+$; calc.: 442.3559), with seven degrees of unsaturation. The ${}^{1}H^-$ and ${}^{13}C^-$ NMR spectra were distinctly similar to those of compound **1**, except for a double bond between C(5) and C(6). Two broad s at δ_H 3.95 and 3.99 were ascribed to H^- C(3) and H^- C(4), respectively. The ${}^{13}C^-$ NMR DEPT spectrum displayed resonances for 28 C-atoms, with seven Me, seven CH₂, nine CH and five quaternary C-atoms. The presence of a C(5)=C(6) bond was also inferred from the HMBC interactions of H^- C(6) (δ_H 5.78) with C(10) (δ_C 39.5), and of C(5) (δ_C 140.5) with H^- C(19) (δ_H 1.04) (Fig. 6). The β -orientation of C(4) -OH was inferred from the $W_{1/2}$ value of H^- C(4) (δ_H 3.99, $W_{1/2} = 5.5$ Hz). Finally, the structure of **5** was deduced as (2'E,20S)-20-(dimethylamino)-4 β -hydroxy-3 β -(tigloylamino)pregn-5-ene.

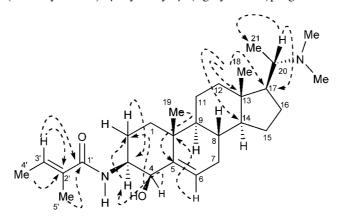


Fig. 6. Selected HMBC interactions in salignarine-F(5)

Salonine-C (6), $C_{28}H_{44}N_2O$, was obtained as a white amorphous solid. Its ¹H-NMR spectrum revealed the presence of 3 s at δ_H 0.86, 1.06, and 1.82, assigned to Me(18), Me(19), and Me(5'), respectively. Me(21) and Me(4') resonated at δ_H 1.73 (d, J =

6.5 Hz) and 1.78 (d, J = 6.9 Hz), respectively, while the Me₂N group resonated at $\delta_{\rm H}$ 2.91 (s). The ¹³C-NMR spectra displayed resonances for 28 C-atoms, with seven Me, six CH₂, eight CH, and seven quaternary C-atoms. In the HMBC spectrum, the olefinic H-atom ($\delta_{\rm H}$ 5.78) showed an interaction with C(3) ($\delta_{\rm H}$ 45.5), which indicated a C(4)=C(5) bond. Another downfield ¹H-NMR signal at $\delta_{\rm H}$ 5.41 showed HMBC interactions with C(16) ($\delta_{\rm C}$ 31.3) and C(17) ($\delta_{\rm C}$ 51.5), and the quaternary C(14)-atom at $\delta_{\rm C}$ 154.6 showed interaction with H–C(18) ($\delta_{\rm H}$ 0.86), indicating the presence of the second C=C bond between C(14) and C(15). Compound **6** was, thus, assigned as (2′E,20S)-20-(dimethylamino)-3 β -(tigloylamino)pregna-4,14-diene.

N-(Formyl(methyl)amino)salonine-B (7), $C_{24}H_{37}NO_2$, was obtained as a yellow gum. The base peak at m/z 356.3233 in the HR-EI mass spectrum was due to the loss of a Me group, which indicated the presence of a C=C bond. The peak at m/z 58 indicated the presence of a formyl(methyl)amino group. The ¹H- and ¹³C-NMR spectra of **7** were distinctly similar to those of compound 3. A m at $\delta_{\rm H}$ 4.19 was ascribed to H-C(20), a split s at $\delta_{\rm H}$ 2.64/2.72 for the MeN group, and a splitted s at $\delta_{\rm H}$ 8.02/8.19 for the formyl H-atom, peak doubling being due to the rotational isomerism of the formamido group. The ¹³C-NMR spectra displayed resonances for 24 C-atoms, with five Me, seven CH₂, eight CH, and four quaternary C-atoms. The β -orientation of the MeO-C(3) group was inferred from a m resonating at $\delta_{\rm H}$ 3.03 ($W_{1/2} = 17.7~{\rm Hz}$). The presence of a C(16)=C(17) bond was supported by the HMBC spectrum (Fig. 7). H-C(16) ($\delta_{\rm H}$ 5.78) showed an interaction with C(15) ($\delta_{\rm C}$ 31.2), and C(17) ($\delta_{\rm C}$ 153.4) with H–C(18) $(\delta_{\rm H}\,0.75)$. The presence of a second double bond between C(5) and C(6) was inferred from the HMBC interactions of H–C(6) ($\delta_{\rm H}$ 5.34) with C(10) ($\delta_{\rm C}$ 35.5), and of C(5) ($\delta_{\rm C}$ 141.1) with H-C(19) ($\delta_{\rm H}$ 1.01). Finally, the structure of compound 7 was deduced as (20S)-20-(formyl(methyl)amino)-3 β -methoxypregna-5,16-diene.

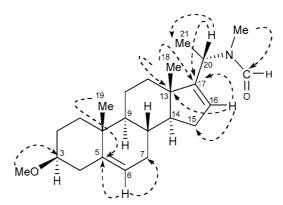


Fig. 7. Selected HMBC interactions in N-(formyl(methyl)amino)salonine-B (7)

The steroidal alkaloid dictyophlebine (8) was isolated for the first time from *S. saligna*. This compound has previously been isolated from *Dictyophleba lucida* [18]. The UV, IR, NMR, and mass spectra of this compound were identical to those reported in the literature. Five more known compounds, epipachysamine-D (9), saracosine (10), iso-*N*-formylchonemorphine (11), sarcodinine (12), and alkaloid-C (13) were also

isolated from S. saligna and structurally identified by comparison with the literature data

The anticholinesterase activities of 1-13 are reported here for the first time. All compounds were found to be nontoxic in brine-shrimp lethality assay ($LD_{50} > 1,000 \text{ mg/ml}$). Table 3 shows their inhibitory activities against the enzymes acetylcholinesterase (AChE) and butyrylcholinesterase (BChE). All these compounds were found to be more selective towards BChE, whereas the standard drug eserine was 21-times more selective towards AChE. Compounds 1-13 were found to be noncompetitive enzyme inhibitors.

Table 3. In vitro Activities (in terms of IC_{50} [μ M] \pm SEM) of Compounds 1–13 against Acetylcholinesterase (AChE) and Butyrylcholinesterase (BChE). For comparison, the respective IC_{50} values for eserine as a reference compound are also shown

Compound	AChE	BChE
1	15.99 ± 0.13	6.91 ± 0.06
2	20.29 ± 1.82	1.89 ± 0.06
3	249 ± 10.23	25.7 ± 0.63
4	82.5 ± 2.22	20.95 ± 3.2
5	30.2 ± 2.0	1.9 ± 0.2
6	7.8 ± 0.5	32.2 ± 0.5
7	48.6 ± 2.7	10.5 ± 0.3
8	6.21 ± 0.23	3.65 ± 0.02
9	28.93 ± 0.54	2.82 ± 0.02
10	20.0 ± 1.30	3.86 ± 0.01
11	6.36 ± 0.22	4.07 ± 0.11
12	40.04 ± 0.13	12.51 ± 0.06
13	42.2 ± 0.26	22.13 ± 0.14
Eserine	0.041 ± 0.00	0.857 ± 0.02

Experimental Part

General. Column chromatography (CC) was carried out on silica gel 60 (70–230 mesh). Pre-coated silicagel 60 F₂₅₄ (Merck; 0.2 mm) glass plates for TLC were inspected at 254 and 366 nm, and developed with Dragendorff's spraying reagent. Melting points (m.p.) were determined in glass capillary tubes using a Büchi-535 melting-point apparatus. Optical rotations were measured on a Schmidt–Haensch Polartronic-D polarimeter. UV Spectra were measured on a Hitachi U-3200 spectrophotometer. The IR spectra were recorded on a Jasco A-302 spectrophotometer. ¹H- and ¹³C-NMR Spectra were recorded on Bruker AM-300 and AMX-500 spectrometers at 300 or 500, and at 75 or 125 MHz, respectively, FAB- and HR-EI-MS: on Jeol JMS-600 and HX-110 mass spectrometers, resp.

Collection and Identification of Plant Material. The whole plants of Sarcococca saligna (D.DON.) Muel. (50 kg) were collected in October 1999 from the District Bagh of Azad Kashmir (Pakistan) and were identified by Mr. Tahir Ali (taxonomist at the Department of Botany, University of Karachi, Pakistan). A voucher specimen (KU*19290) was deposited in the Herbarium of the University of Karachi.

Extraction. The air-dried plant material (14 kg) was crushed and soaked in MeOH (50 l) for 15 d. The MeOH extract was filtered and evaporated to a gum (1.25 kg), triturated in distilled H_2O (5 l), and subsequently extracted with petroleum ether (251 g, Fraction A); CHCl₃ at pH 3 (6.0 g; Fraction B), pH 7 (220 g; Fraction C) and pH 9 (25.0 g; Fraction D); AcOEt (95 g; Fraction E); and, finally, with BuOH (145 g; Fraction F). Fractions B and D showed 100% inhibition against acetylcholinesterase and butyrylcholinesterase (25 mg/ml). These two fractions were subjected to repeated CC (1.5 kg SiO₂), eluting with petroleum ether/acetone/Et₂NH mixtures of increasing polarity. The resulting subfractions were individually subjected to further CC on SiO₂

(petroleum ether/acetone/Et₂NH mixtures) to obtain semipure sub-subfractions, which were finally purified by prep. TLC (SiO₂; solvent mixtures as above) to obtain pure 1-13.

 $\begin{array}{l} 2\text{-}Hydroxysalignarine-E \ (= (2'\text{E},20\text{S})\text{-}20\text{-}(dimethylamino)\text{-}2\beta\text{-}hydroxy\text{-}3\beta\text{-}(tigloylamino)pregn-}14\text{-}ene; \ \textbf{1}). \\ \text{Yellow gum. Yield: 19.7 mg (141 ppm). } [a]_{D}^{20} = +38.9 \ (c = 0.3, \text{MeOH}). \ \text{UV (MeOH): 211 (1.32), 229 (2.76). IR (CHCl_3): 3329 (NH, OH), 2911 (CH), 1663 (C=O), 1624 (C=C). }^{1}\text{H-} \ \text{and } ^{13}\text{C-NMR: see } \textit{Tables } 1 \ \text{and } 2, \text{ resp.} \\ \text{EI-MS: 442 (34) } (M^+), 427 \ (100, [M-15]^+), 328 \ (2), 296 \ (5), 100 \ (9), 83 \ (17), 72 \ (39), 55 \ (7). \ \text{FD-MS: 442 (C$_{28}$H$_{46}$N$_{2}$O$_{2}). } \\ \text{HR-EI-MS: 442.3489 } (M^+, \ C$_{28}$H$_{46}$N$_{2}$O$_{2}^+; calc.: 442.3559), 83.0504 \ (C$_{3}$H$_{7}$O$^+; calc.: 83.0496), 72.0803 \ (C$_{4}$H$_{10}$N$^+; calc.: 72.0813). \\ \end{array}$

5,6-Dihydrosarconidine (=(20S)-20-(dimethylamino)-3β-(methylamino)-5α-pregn-16-ene; **2**). Colorless powder. Yield: 21.03 mg (150 ppm). $[\alpha]_D^{30} = -60$ (c = 0.2, CHCl₃). UV (MeOH): 212 (2.66). IR (CHCl₃): 3650 (NH), 2810 (CH), 1601 (C=C). 1 H- and 1 3C-NMR: see *Tables 1* and 2, resp. EI-MS: 358 (3) (M^+), 343 (100, $[M-15]^+$), 72 (47), 58 (14). FD-MS: 358 ($C_{24}H_{42}N_2^+$). HR-EI-MS: 358.3193 (M^+ , $C_{24}H_{42}N_2$; calc.: 358.3198), 72.0811 ($C_{4}H_{10}N^+$; calc.: 72.0813).

Salignamine (= (20S)-20-(methylamino)-3β-methoxypregn-5,16-diene; **3**). Yellow gum. Yield: Yellow gum. Yield: 9.9 mg, (70.7 ppm) $[a]_D^{10} = -23$ (c = 0.12, CHCl₃). UV (MeOH): 202 (2.9). IR (CHCl₃): 3303 (NH), 2901 (CH), 1386 (C-O). 1 H- and 1 C-NMR: see *Tables 1* and 2, resp. EI-MS: 343 (16, M^+), 328 (100, $[M-15]^+$; 312 (14), 58 (37). FD-MS: 343 (C_{23} H₃₇NO $^+$). HR-EI-MS: 343.2880 (M^+ , C_{23} H₃₇NO; calc.: 343.2875).

2-Hydroxysalignamine (=(20S)-20-(dimethylamino)-2 β -hydroxy-3 β -methoxypregn-5,16-diene; **4**). Yellow gum. Yield: 6.9 mg, (44.7 ppm). [α] $_{20}^{10}$ = -26 (c = 0.02, CHCl₃). UV (MeOH): 206 (3.8). IR (CHCl₃): 2905 (CH), 1663 (C=O). 1 H- and 13 C-NMR: see *Tables 1* and 2, resp. EI-MS: 373 (34, M^+), 358 (100, [M – 15] $^+$, 342 (2), 72 (39). FD-MS: 373 (C_{24} H₃₉NO $_{2}^+$). HR-EI-MS: 373.2982 (C_{24} H₃₉NO $_{2}^+$; calc.: 373.2980), 72.0803 (C_{4} H₁₀N $^+$; calc.: 72.0813).

Salignarine-F (= (2′E,20S)-20-(dimethylamino)-4β-hydroxy-3β-(tigloylamino)pregn-5-ene; **5**). Yellow gum. Yield: 6.79 mg, (40.3 ppm). $[a]_{10}^{20} = -71$ (c = 0.014, MeOH). UV (MeOH): 203 (2.6). IR (KBr): 3650 (NH), 2810 (=CH), 1601 (C=C), 1511 (NH). 1 H- and 13 C-NMR: see *Tables 1* and 2, resp. EI-MS: 442 (3, M^{+}), 427 (70, $[M-15]^{+}$), 84 (37), 72 (100), 58 (14). FD-MS: 442 (C_{28} H₄₆N₂O₂ $_{2}$). HR-EI-MS: 442.3577 (M^{+} , C_{28} H₄₆N₂O₂ $_{2}$; calc.: 442.3559), 427.3233 (C_{27} H₄₃N₂O₂ $_{2}$; calc.: 427.3273), 83.0503 (C_{5} H₇O+; calc.: 83.0496), 72.0811 (C_{4} H₁₀N+; calc.: 72.0813).

Salonine-C (= (2′E,20S)-20-(dimethylamino)-3β-(tigloylamino)pregna-4,14-diene; **6**). White amorphous solid. Yield: 20.03 mg, (140 ppm). [α]₀²⁰ = -120 (c = 0.048, MeOH). UV (MeOH): 206 (2.23). IR (CHCl₃): 3650 (NH), 2810 (=CH), 1601 (C=C), 1511 (NH). ¹H- and ¹³C-NMR: see *Tables 1* and 2, resp. EI-MS: 424 (3, M⁺), 409 (25, [M – 15]⁺, 84 (37), 72 (100), 58 (14). FD-MS: 424 (C₂₈H₄₄N₂O⁺). HR-EI-MS: 424.3489 (M⁺, C₂₈H₄₄N₂O⁺; calc.: 424.3508), 409.3233 (C₂₇H₄₁N₂O⁺; calc.: 409.3273), 83.0503 (C₅H₇O⁺; calc.: 83.0496), 72.0811 (C₄H₁₀N⁺; calc.: 72.0813).

N-[Formyl(methyl)amino]salonine-B (=(20S)-20-[formyl(methyl)amino]-3 β -methoxypregna-5,16-diene (7). Yellow gum. Yield: 5.03 mg, (15.0 ppm). [a] $_0^2$ 0 = -21 (c=0.075, MeOH). UV (MeOH): 237 (2.86). IR (CHCl $_3$): 2810 (=CH), 1601 (C=C). 1 H- and 1 C-NMR: see *Tables 1* and 2, resp. EI-MS: 371 (3, M^+), 356 (100, [M-15] $^+$), 58 (14). FD-MS: 371 (C_2 4H $_3$ 7NO $_2^+$). HR-EI-MS: 371.2982 (M^+ , C_2 4H $_3$ 7NO $_2^+$; calc.: 371.2980), 356.3233 (C_2 3H $_3$ 4NO $_2^+$; calc.: 356.3273).

Inhibition Assays. Electric eel acetylcholinesterase (AChE), horse butyrylcholinesterase (BChE), acetylthiocholine iodide, butyrylcholine chloride, 5.5'-dithiobis(2-nitrobenzoic acid) (DTNB; =5-[(3-carboxy-4-nitrophenyl)disulfanyl]-2-nitrobenzoic acid), and eserine were purchased from Sigma (St. Louis, MO). Buffer and other chemicals were of analytical grade. AChE Inhibition was determined spectrophotometrically, with acetylthiocholine as substrate, by modifying the method of Ellman [16]. The reaction was carried out in $100~\mu m$ sodium phosphate buffer (pH 8.0) at 25° . In a typical assay, $140~\mu l$ of buffer, $20~\mu l$ of enzyme preparation, and $20~\mu l$ of test-compound soln. were mixed and incubated for 30~min. DTNB ($10~\mu l$) was added, and the reaction was initiated by adding $10~\mu l$ of acetylthiocholine. Butyrylthiocholine chloride was used as a substrate to assay BChE under similar conditions as above. The rate of hydrolyses of acetylthiocholine and butyrylthiocholine were determined by monitoring the formation of the yellow 2-nitro-5-sulfanylbenzoate anion (as a result of the reaction of DTNB with the thiocholine released by the enzymatic hydrolysis) at a wavelength of 412 nm.

This work was supported by the *Pak-Kazakh* joint-research program of the *Ministry of Science and Technology*, Pakistan. *Z.-u. H.* gratefully acknowledges the financial support of Mr. *Sheikh Ahmed Bombal*, President of the *Faran Club International*. Financial support by Mr. *Ateeq-ur-Rahman Barry* (*M/S Haseen Habib Trading Co.*) to *S. A. N.* is also gratefully acknowledged.

REFERENCES

- [1] W. Sippl, J. M. Contreras, Y. Rival, C. G. Wermuth, in 'Rational Approaches in Drug Design', Eds. H.-D. Höltje, W. Sippl, Prous Science Press, Barcelona, 2001, p. 56-64.
- [2] E. Nasir, S. I. Ali, 'The Flora of West Pakistan', Fakhri Printing Press, Karachi, 1974, 65, 2.
- [3] M. Kiamuddin, H. K. M. A. Hye, Pak. J. Sci. Ind. Res. 1970, 13, 59.
- [4] Atta-ur-Rahman, M. R. Khan, M. I. Choudhary, M. Z. Iqbal, Phytochemistry 1997, 45, 861.
- [5] J. M. Kholi, A. Zaman, A. R. Kidwai, Tetrahedron 1967, 23, 3829.
- [6] J. M. Kholi, A. Zaman, A. R. Kidwai, Phytochemistry 1971, 10, 442.
- [7] U. L. B. Jayasinghe, M. Nadeem, Atta-ur-Rahman, M. I. Choudhary, H. D. Ratnayake, Z. Amtul, Nat. Prod. Lett. 1998, 12, 103.
- [8] I. Naeem, N. Khan, M. I. Choudhary, Atta-ur-Rahman, Phytochemistry 1996, 43, 903.
- [9] R. Goutarel, C. Conreur, L. Djakouré, M. Leboeuf, A. Cavé, Tetrahedron 1968, 24, 7013.
- [10] Z.-M. Zou, L.-J. Li, M. Yang, S.-S. Yu, P.-Z. Cong, D.-Q. Yu, *Phytochemistry* **1997**, 46, 1091.
- [11] Atta-ur-Rahman, S. Anjum, A. Farooq, M. R. Khan, M. I. Choudhary, Nat. Prod. Lett. 1998, 11, 297.
- [12] Atta-ur-Rahman, M. I. Choudhary, M. R. Khan, M. Z. Iqbal, Nat. Prod. Lett. 1998, 11, 81.
- [13] M. Chiu, R. Nie, Z. Li, J. Zhou, J. Nat. Prod. 1992, 55, 25.
- [14] Atta-ur-Rahman, S. Anjum, A. Farooq, M. R. Khan, Z. Parveen, M. I. Choudhary, J. Nat. Prod. 1998, 61, 202.
- [15] Atta-ur-Rahman, Zaheer-ul-Haq, A. Khalid, S. Anjum, M. R. Khan, M. I. Choudhary, Helv. Chim. Acta 2002, 85, 678
- [16] G. L. Ellman, D. K. Courtney, V. Andres, R. M. Featherstone, Biochem. Pharmacol. 1961, 7, 88.
- [17] H. Budzkiewicz, C. Djerassi, D.H. Williams, 'Structure Elucidation of Natural Products by Mass Spectrometry', Holden-Day, New York, 1964, 2, 5.
- [18] K.-H. Qui, X. Monseur, M. T. Ho, R. Kocjan, R. Goutarel, Bull. Soc. Chim. Fr. 1965, 10, 3035.

Received August 26, 2003